#### **REVIEW ARTICLE**



# Structure–activity relationship of polyamine conjugates for uptake via polyamine transport system

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#### Abstract

High toxicity of anticancer drugs led to development of targeted drug delivery directly to the specific organs. Polyamine transport system (PTS) of mammalian cells is one of the targets for a cell-selective drug delivery of polyamine–drug conjugates into specific organs. Even without having a 3D structure for mammalian PTS, synthesis of polyamine derivatives and evaluation of their cytotoxic effects are potential practical approaches to find optimal polyamine moieties to be transported from the PTSs. Chinese hamster ovary (CHO) and its mutant cell line (CHO-MG) are two important cells for evaluation of polyamine transporters (PAT). If a polyamine conjugate ligand demonstrates a high IC<sub>50</sub> ratio on CHO-MG/CHO cells, this indicates a high selectivity of such compound toward PAT. This study discussed the structural requirements (charge, linker, vector, cargo) of polyamine conjugates in order to be transported into the cells by the mean of PTS.

**Keywords** Polyamine conjugates  $\cdot$  Polyamine transport system (PTS)  $\cdot$  Membrane transporter  $\cdot$  Chinese hamster ovary (CHO)  $\cdot$  CHO-MG

## Introduction

The polyamines (PAs), including spermine, spermidine, and putrescine, are an essential class of metabolites with flexible aliphatic carbon chains. Although PAs have simple structure, yet they are essential for eukaryotic cells growth [1–3]. Spermine was discovered by Leuwenhoek in 1678 in seminal fluid [4, 5]. These polycations are bonded to anions by reversible ionic interactions to DNA, RNA, proteins, phospholipids, and mediate molecular functions such as maintenance of chromatin conformation, regulation of specific gene expression, regulation of ion-channel, the stability of membrane, and free-radical scavenging [6–10]. During the last decades, the metabolism of polyamines is studied as a potential target for cancer treatment.

The first rate-limiting step in polyamine biosynthesis is production of putrescine, which is tightly regulated by ornithine decarboxylase (ODC), a phosphate-dependent

Lotfollah Saghaie saghaie@pharm.mui.ac.ir decarboxylase. It has a short half-life and is controlled at several steps including transcription, post-transcriptional processing, and changes in translational efficiency and altered stability of the protein [11-13] ODC is inhibited by 2-difluoromethylornithine (DMFO) that decrease cellular ODC enzyme activity [14]. Another rate-limiting enzyme in polyamine synthesis can be S-adenosylmethionine decarboxylase (AdoMetDC) that produces the aminopropyl donor via the aminopropyl transferases spermidine synthase and spermine synthase, respectively for the synthesis of both spermidine and spermine [15–17].

Moreover, the intracellular amounts of polyamine are adjusted through the activity of two enzymes involved in polyamine catabolism. Spermidine/spermine N1-acetyltransferase (SSAT) is the first catabolic enzyme of PAs which transfer an acetyl group to the N position of spermine or spermidine, from an acetyl-coenzyme A molecule, and produce N1acetylspermine or N1-acetylspermidine, respectively [18, 19]. The acetylated polyamines then can either be transferred outside the cell by diamine exporter (DAX) [20]or be used as a substrate for flavin-dependent polyamine oxidase (APAO), in which it converts the N1-acetylspermine to spermidine and N1-acetylspermidine to putrescine [21, 22].

Polyamine transport system is more studied in prokaryotic system rather than eukaryotic systems. In eukaryotic cells, this

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transport system plays an essential role in maintaining intracellular polyamine homeostasis via a known energydependent and carrier-mediated polyamine uptake mechanism. The first model was described by Poulin et al. and addressed the structural characterization of mammalian polyamine transport system [23]. They suggested that polyamines are first entered into the cell by a membrane carrier followed by localization of PAs in polyamine-sequestering vesicles (PSVs) through a mechanism that requires a directed pH gradient and proton exchange. The second model was described by Belting et al. (1999, 2003), suggesting that before being transported to the cell, spermine is binds to heparan sulfate groups of GPC1 heparan sulfate on the cell surface, and when it is inside the cell, the spermine is released by nitric oxide synthase-2 (NOS2) activity through NO-oxidation-mediated process [24, 25].

Chemotherapeutic compounds are highly toxic, and serious side effects are inevitable consequences of treatment with these compounds; thus, specific targeting and delivery of therapeutic molecules into a certain organ, tissue, or cell have been progressively studied as alternative treatment approaches, particularly for cancer. On the one hand, previous reports demonstrated that in tumor cells, polyamine metabolisms, including biosynthesis and uptake, are particularly amplified, and the efficiency of the polyamine transport system (PTS) is significantly increased [26]. This particular characteristic of tumoral cells allows for cell-selective drug delivery; in other words, targeted delivery of polyamine–drug conjugates into specific cell types is a possibility [27].

On the other hand, the PTS system in mammalian cells is poorly understood [11] and the majority of the studies have been addressing the therapeutic effects of homologous derivatives of polyamines. The effective factors in designing PAs derivatives are (i) increasing the length of polyamine backbone, (ii) the atom type responsible for the attachment of polyamine to the drug, and (iii) the physiochemical properties of the drugs.

Chinese hamster ovary (CHO) with a mutant cell line (CHO-MG) is an important mean for detection and screening the selective conjugate delivery by polyamine transporters (PAT) [28]. For example, MGBG methylglyoxal bis-(guanylhydrazone) is a PA-conjugated drug and its transportation is PTA-mediated. It has a high toxicity for CHO, while CHO-MG cell line is much more resistant to this drug [29]. Therefore, determination and comparison of the half maximal inhibitory concentration (IC<sub>50</sub>) of such drugs (PAs with PTS-mediated transport systems) using these two CHO lines can be used to estimate the delivery rate of the drugs by PAT [30]. In other words, to obtain polyamine conjugate ligands with high selectivity toward the PTS system, achieving a high (CHO-MG/CHO) IC<sub>50</sub> ratio in in vitro studies is a must [31].

This review study aimed at addressing important structural factors for PTS transportation of polyamine-derivative drug



**Fig. 1** The structure of deferiprone (compound 1) and hydroxypyridinone–polyamine conjugate (compound 2)

conjugates, including charge, linker, and vector length cargo effects.

# Structure-activity research of drug-polyamine conjugates

### **Charge effects**

Iron chelators and their chemical charge are of critical importance for transportation of PAs [32–34]. Ligand donors in iron chelators, e.g., carboxylates, are highly polar and often ionizable. The transporter tendency is largely toward binding to neutral or positively charged ligands rather than negatively charged ones.

Bergeron et al. [35] synthesized 1-(12-amino-4,9diazadodecyl)-2-methyl-3-hydroxy-4(1H)-pyridinone (Fig. 1, compound 2), a bidentate hydroxypyridinone–polyamine conjugate, and evaluated its IC<sub>50</sub> against L1210 murine leukemia cell line in vitro at a concentration of 0.2 M and reported that compared to the parent ligand (Fig. 1, compound 1), the conjugated drug demonstrated > 230 folds increase in its effect.

Bergeron et al. synthesized polyamine conjugates using neutral chelator (L1 conjugate) [35] with a PAT-mediated transportation mechanism. (S)-2-(2,4-Dihydroxyphenyl)-4,5dihydro-4-methyl-4-thiazolecarboxylic acid [(S)-4'-(HO)-



(S)-4-(HO)-DADFT (3)



Fig. 2 The structure of iron chelator (compound 3) and polyamine chelator conjugate (compounds 4 and 5)



**Fig. 3** The structure of a new generation of iron chelators designed based on 8-hydroxyquinoline (8-HQ) scaffold which are linked to linear polyamine

DADFT] (Fig. 2, compound 3) is an active oral analogue of desferrithiocin (DFT). DFT itself is a neutral iron chelator isolated from *Streptomyces* antibiotics [36].

Later, Bergeron et al. (2005, 2010) [37, 38] synthesized (S)-2-(2,4-Dihydroxyphenyl)-4,5-dihydro-4-methyl-4thiazolecarboxylic acid [(S)-4'-(HO)-DADFT] (Fig. 2, compound 4) that was attached to norspermidine (NSPD) at the C-4'-oxygen through a butyl group and its ethyl ester analogue (NSPD-(S)-4'-(HO)-DADFT-EE) (Fig. 2, compound 5). The IC<sub>50</sub> value (IC<sub>50</sub> = 40  $\mu$ M) of acid conjugates (4) was more than the ester analogue (5) (IC<sub>50</sub> = 1.5  $\mu$ M); thus, antiproliferative effects of (4) are less than the ester analogue. Moreover, as the name suggests, the (S)-4'-(HO)-DADFT acid analogue (Fig. 2, compound 4) has an acidic functional group at its chelator fragment, which has negatively charge at physiological pH (i.e., carboxylates). This negative charge is hindrance for transformation by PAT and is not likely to be taken up by PAT cells.

### Linker effects

Corcé et al. synthesized a new generation of iron chelators, quilamines, which are based on 8-hydroxyquinoline (8-HQ) scaffold linked to linear polyamine vectors [31]. The synthesized quilamines were differed in length (HQ0-44 (6), HQ1-44 (7), HQ2-44 (Fig. 3, compound 8)) and/or the chemical

**Fig. 4** A model of the structural constraints of the PTS proposed by Phanstiel [41] and the transporter pathway which was modified according to Padariya et al. [42]

nature of the spacer compound, i.e., HQCO-44 (Fig. 3, compound 9) and HQCS-44 (Fig. 3, compound 10)) (Fig. 3). The difference between the added ligands of compounds 9 and 10 is their heteroatom containing either amide or thioamide functional group, respectively. The IC<sub>50</sub> ratio of these compounds against CHO-MG/CHO was assessed, which as mentioned above, is a mean to measure the selectivity for PTS. According to the IC<sub>50</sub> ratio, those ligands differ in the length of the spacer and showed different IC<sub>50</sub> ratio values (different selectivity toward PTS) and higher antiproliferative activities (IC<sub>50</sub> values between 0.4 and 1.8  $\mu$ M) while ligands differ in the chemical nature of the spacer (9, 10) were less efficient (IC<sub>50</sub> values 35.25  $\mu$ M), with lower IC<sub>50</sub> ratio values.

Corcé et al. design [31] was based on a structural constraints model of the PTS proposed by Phanstiel and Delcros (Fig. 4) [39–41], in which they indicated that the charge and size of polyamines are critical parameters for synthesis of PTS-targeting compounds.

According to this model, insertion of carbonyl and thiocarbonyl groups between iron chelator and polyamine chain resulted in decreased affinity of quilamines **9** and **10** toward PTS, in a sense that the (thio)amide group inhibited protonation of the nitrogen at physiological pH, omitting the electrostatic interaction required for recognition and attachment to the PTS (Fig. 5).

The quilamine compounds which were wary based on their length (6, 7, 8) have three nitrogen atoms which can form three positive charges at physiological pH, an important feature that requires for higher affinity toward PTS and transportation by this system [39, 40], and as a result of higher PTS selectivity, these compounds have higher  $IC_{50}$  ratio as well. The length of the spacer between N1 nitrogen and the chelator plays an essential role in PAT selectivity as well. Corcé et al. suggested that the presence of a hydrophobic pocket within the recognition site of the PTS favors the affinity (Fig. 6) [31]. Moreover, compared to quilamine compounds 6 and 7, increasing the size of the spacer in compound 8 led to a better



Fig. 5 A hypothetical model, based on Phanstiel and Delcros [41], to explain selectivity loss of quilamine compounds 9 and 10 toward PTS



fit the chelator moiety inside the hydrophobic cavity, resulting a better hydrophobic interaction, in other words a stronger affinity/selectivity toward PTS (Fig. 6).

An interesting finding was the higher  $IC_{50}$  ratio of quilamine **6** than that of quilamine **7**. Corcé et al. suggested that in the case of N1 nitrogen, which is cationic at the physiological pH, its positive charge is distributed between the N1 nitrogen itself and the nitrogen of the heteroaromatic ring. Therefore, this delocalization resulted in a higher electrostatic interaction and as a result an increased PTS selectivity (Fig. 7).

The linker effects were studied using two groups of compounds. Phanstiel et al. [41] studied the first group in which have 1-naphthyl moiety. Some derivatives had a methylene (compound **10**) or ethylene analogue (compound **11**) linkers which showed a high selectivity toward PAT with an IC<sub>50</sub> ratios > 164, but the propylene analogue (compound **12**) showed no PAT selectivity (IC<sub>50</sub> ratios: no activity) (Fig. 8).

The second group was studied by Breitbeil et al. in which the polyamine was conjugated with anthracenyl moiety (Fig. 9). Increasing the linker length from methylene (compound 13) with a CHO  $IC_{50}$  of 0.45 µM to ethylene analogue (compound 14) with a CHO  $IC_{50}$  of 9.8 µM and propylene (compound 15) with a CHO  $IC_{50}$  of 130.1 µM) changed the  $IC_{50}$  ratio, in which their CHO-MG/CHO  $IC_{50}$ ratios were calculated at 148, 3.4, and 1, respectively [43]. Increasing the length of the linkers leads to formation of other conformations or molecular shapes, which moved the bulky aryl group out of the hydrophobic pocket, which itself resulted in a decreased PAT selectivity. In other words, there is a significant decrease in drug uptake via the PAT when the linker length was increased.

#### **Cargo effects**

Phanstiel et al. [44] synthesized several N1-substituted polyamines containing various cargo units (Fig. 10) including benzyl (compound 16), naphthyl (compound 17), anthracenyl (compound 18), and pyrenyl (compound 19). PAT selectivity of these compounds was studied by evaluating their IC<sub>50</sub> against CHO-MG/CHO. In comparison with compound 17 with a (CHO-MG/CHO) IC<sub>50</sub> ratio more than 164, the

**Fig. 6** The hypothetical model of the structure–selectivity relationship between compound **8** and PTS according to Phanstiel and Delcros model [41]



**Fig. 7** The hypothetical model of the structure–selectivity relationship between compound **6** and PTS according to Phanstiel and Delcros model [41]





reported IC<sub>50</sub> ratio for compound **16** was not significant. It is suggested that the bulky N-aryl substituent increased the binding affinity toward PAT, and as a result, the cytotoxicity of the polyamine conjugates in vitro. However, on the other hand, increasing the size of the arenyl unit from that of compound **17** to a more hydrophobic and bulkier cargo unit such as the ones in compounds **18** and **19** resulted in lower selectivity for the polyamine transporter. The respective CHO-MG/CHO IC<sub>50</sub> ratios for compounds **18** and **19** conjugates were 148 and 34, respectively. Therefore, the hydrophobicity and the size of the arenyl unit are significant influencing parameters for the PAT selectivity profile of conjugates, in a sense that the polyamines conjugated with a compound that its cargos have moderate hydrophobicity and size (e.g., naphthyl) demonstrated higher (CHO-MG/CHO) IC<sub>50</sub> ratios.

Phanstiel et al. [45] synthesized several mono, di, and trisubstituted arylene–polyamine conjugates (Fig. 11) and investigated their PAT-mediated transportation yield inside the cell. They synthesized three disubstituted compounds (anthryl, naphthyl, benzyl) which demonstrated the highest reported CHO-MG/CHO IC<sub>50</sub> ratios among all the related studies with a value of > 2222, > 833, and 677, respectively. It is suggested that these disubstituted derivatives have an extremely high selectivity toward PAT of the CHO cells. There is a direct correlation between the size of arylene groups of the disubstituted derivatives and the related PAT selectivity. On the one hand, increasing the size of anthracenyl units resulted in a higher affinity/selectivity toward PAT. In contrast, in monosubstituted increasing, the size of the aryl unit resulted in a lower affinity/selectivity toward PAT [45]. The selectivity of the trisubstituted compound (**25**) was not determined because CHO and CHO-MG IC<sub>50</sub> were > 500 and IC<sub>50</sub> ratio was not applicable.

One of the most serious issues arising from using polyamine-based derivatives is the metabolism by polyamine oxidase (PAO), in which by degrading the compound it









significantly reduces the PTS selectivity of a polyamine–drug conjugates. To tackle this issue in a cell culture, the aminoguanidine (as a known inhibitor of PAO) is added to the medium and prevents the degradation of such compounds. An alternative approach to handle PAO issue is N-methylation of primary amines [46].

Phanstiel et al. [47] combined N-methylation method with the synthesis of disubstituted derivatives to obtain superior, more metabolically stable PTS ligands. The primary experiment was performed with and without aminoguanidine (AG) and using nonmethylated analogues, in which the absence of AG resulted in a dramatic decrease in PTS selectivity, for instance, CHO-MG/CHO IC<sub>50</sub> ratio of compound **26** > 3571 and 2.1, with and without AG, respectively. Absence of AG additive resulted in a significantly lower PTS selectivity for compounds **28** and **30**, while with AG, they resulted in IC<sub>50</sub> ratio of > 4645 and 727, respectively.

The second experiment was performed with and without aminoguanidine (AG) and using methylated compounds (27, 29, 31). Using N-methylation strategy resulted in a higher metabolic stability in which the CHO-MG/CHO IC<sub>50</sub> ratio with AG for compound 27 was > 1204, while without the AG, it was measured at > 1190. Ultimately, the disubstituted platforms, which enhanced PTS targeting, and the N-methylation strategies, which increase the metabolic stability,

are two effective approaches in designing compounds with desirable affinity/selectivity toward the PTS system (Fig. 12).

### **Vector lengths**

In several studies, N1-substituted tri- and tetraamines with different tether lengths (number of CH2 spacer units) between nitrogen centers were synthesized (Fig. 13) [39, 40, 48]. Biological evaluation of different vectors (triamines and tetraamine motif) using CHO cell lines revealed that all polyamine vectors do not have the same selectivity, and only a few of the compounds demonstrated a desirable PAT selectivity characteristics. A comparison between compound **41** and **44** (Fig. 13) suggests that using compound **44** with longer tether leads to a significant increase in polyamine uptake with a CHO-MG/CHO IC<sub>50</sub> ratio of 148. However, using all tetraamine conjugates resulted in a dramatic decrease in PAT selectivity ( $0.7 < IC_{50}$  ratio < 3.1).

By taking into account the  $IC_{50}$  ratio and the vector design, triamine derivatives (compounds **41–46**) showed a significantly higher PTS selectivity compared to the tetraamine conjugates (compounds **32–40**). It is reported that 4,4-triamine motif (homospermidine) was the optimal chemical structure to increase the PTS-dependent cell entry.

**Fig. 11** The structure of mono, di, and trisubstituted derivatives of arylene-polyamine analogue



Fig. 12 The structure of disubstituted derivatives of polyamine analogue with different moieties



31: R=Me, 6HCI (MeN44Bn44NMe

Gaboriau et al. [49] developed new polyaminoquinoline iron chelators with different polyamine vectors (Fig. 14). In spite of different cellular targets of these quilamines and polyamine–anthracene conjugates (compounds 41, 42, and 50) (iron depletion vs. DNA intercalation), there was a good agreement between two conjugates. Based on the IC<sub>50</sub> of these compounds, triamine analogues resulted in higher PTS selectivity, but it was decreased in the case of 4,4,4-tetraamine. The observed IC<sub>50</sub> for compounds 52 and 47 was 249 and 38 while for 55 and 51 it was 21 and 18, respectively. These findings suggest that quilamines with a 4,4 polyamine chain (compounds **52** and **47**) shows higher PTS selectivity than those of 3,3 chains (compounds **51** and **55**). Moreover, the polyamine conjugates bearing 3,4 chains (**54**, **48**, **50**; with  $IC_{50}$  ratio of 2, 4, and 4, respectively) were wither weakly transported or were not transported by PTS.

Phanstiel et al. reported that the substitution degree at the N1 position in the polyamine vectors (Fig. 15) is a critical factor in the selective delivery of polyamines via the PAT [50]. The N1 anthracenylmethylhomospermidine (compound

Fig. 13 The structure of anthracenyl–polyamine conjugates with different tether lengths



**32**: m=1, n=1, o=2 **35**: m=1, n=3, o=2 **38**: m=3, n=2, o=1 **33**: m=1, n=2, o=1 **36**: m=2, n=2, o=1 **39**: m=3, n=2, o=2 **34**: m=1, n=2, o=1 **37**: m=2, n=2, o=2 **40**: m=3, n=3, o=2





Fig. 14 The structure of polyaminoquinoline iron chelators with different tether lengths

**57**) which contains a secondary nitrogen at the N1 position showed a CHO-MG/CHO  $IC_{50}$  ratio of 148, and compound **56** with a tertiary nitrogen at the N1 position showed a CHO-MG/CHO  $IC_{50}$  ratio of 1. Moreover, PAT selectivity test of the 3°-amine-containing derivatives at the N1 position of dihydroMotu (3,3) and dihydroMotu (4,4) showed a CHO-MG/CHO  $IC_{50}$  ratio of 1; in other words, these compounds were not PAT-selective. Therefore, compounds containing an N1 tertiary substitution pattern are not selective for PAT.

### Conclusion

Although the molecular structure of mammalian PTS is poorly understood, the structure–activity relationships of polyaminelike molecules which are transported by this system have been an interesting field of research particularly for the PTS targeting anticancer drug delivery treatment approaches. With regard to the unclear 3D structure of the PTS, indirect approaches were developed and have been used to study the transport of polyamine conjugates using the PTS in mammalian cells, in which using CHO-MG and CHO cell, two PTSdeficient cell lines with different susceptibilities toward PTStargeted drugs, is one of the most important developed techniques in this regard. The CHO-MG cell line was particularly selected due to its resistance against MGBG methylglyoxal bis-(guanylhydrazone); thus, while polyamine conjugate drugs with high selectivity toward the PTS can impose highly toxic effects on the CHO cells, it demonstrates lower toxicity on CHO-MG cells. This is an important aspect for two reasons, first, compare to native cells, the PTS-deficient cells have lower polyamines uptakes, and second, the CHO-MG/CHO IC<sub>50</sub> ratio determination in these two CHO lines provided a relative delivery yield using PAT. Moreover, a high IC<sub>50</sub> ratio is an important factor for screening of highly selective PTS polyamine conjugate ligands. In this review, we discussed the structure–delivery relationship of polyamines for PTS targeting with regard to the effects of their charge, linker, vector length, and cargo.

In terms of charge, compared to negatively charged conjugates, the neutral or positively charged ligands are much favorable to bind with the PTS. Moreover, increasing the length of the linkers resulted in decreased drug uptake by PAT. Furthermore, with regard to the vector length of the conjugates, 4,4-triamine motif (homospermidine) was found to be the optimal chemical structure with an increased cell entry using PAT. Finally, the cargo effect was evaluated using hydrophobicity and the size of arenyl unit, and it is shown that cargos with moderate hydrophobicity and size have a higher uptake ratio by PAT. In conclusion, to design a successful PTS-targeting polyamine-conjugated drug, many parameters need to be considered.





#### Compliance with ethical standards

**Conflict of interest** The authors declare that they have no conflict of interest.

# References

- 1. Gerner EW, Meyskens Jr FL (2004) Polyamines and cancer: old molecules, new understanding. Nat Rev Cancer 4(10):781
- 2. Cohen SS (1998) Guide to the polyamines, 1st edn. Oxford University Press, London
- Marton LJ, Pegg AE (1995) Polyamines as targets for therapeutic intervention. Annu Rev Pharmacol Toxicol 35(1):55–91
- TABOR H, Tabor CW (1964) Spermidine, spermine, and related amines. Pharmacol Rev 16(3):245–300
- Bachrach U (2010) The early history of polyamine research. Plant Physiol Biochem 48(7):490–495
- Sakai TT, Torget R, Freda CE, Cohen SS (1975) The binding of polyamines and of ethidium bromide to tRNA. Nucleic Acids Res 2(7):1005–1022
- Feuerstein BG, Williams LD, Basu HS, Marton LJ (1991) Implications and concepts of polyamine-nucleic acid interactions. J Cell Biochem 46(1):37–47
- Wang J-Y, Casero RA (2006) Polyamine cell signaling: physiology, pharmacology, and cancer research, 1st edn. Springer-Verlag New York, LLC
- Ha HC, Sirisoma NS, Kuppusamy P, Zweier JL, Woster PM, Casero RA (1998) The natural polyamine spermine functions directly as a free radical scavenger. Proc Natl Acad Sci 95(19): 11140–11145
- Kurata HT, Marton LJ, Nichols CG (2006) The polyamine binding site in inward rectifier K+ channels. J Gen Physiol 127(5):467–480
- Poulin R, Casero R, Soulet D (2012) Recent advances in the molecular biology of metazoan polyamine transport. Amino Acids 42(2–3):711–723
- Shantz LM (2004) Transcriptional and translational control of ornithine decarboxylase during Ras transformation. Biochem J 377(1): 257–264
- Pegg AE (2006) Regulation of ornithine decarboxylase. J Biol Chem 281(21):14529–14532
- Nowotarski SL, Shantz LM (2010) Cytoplasmic accumulation of the RNA-binding protein HuR stabilizes the ornithine decarboxylase transcript in a murine nonmelanoma skin cancer model. J Biol Chem 285(41):31885–31894
- Ikeguchi Y, Bewley MC, Pegg AE (2006) Aminopropyltransferases: function, structure and genetics. J Biochem 139(1):1–9
- Korhonen V-P, Halmekytö M, Kauppinen L, Myöhänen S, Wahlfors J, Keinänen T, Hyvönen T, Alhonen L, Eloranta T, Jänne J (1995) Molecular cloning of a cDNA encoding human spermine synthase. DNA Cell Biol 14(10):841–847
- Wahlfors J, Alhonen L, Kauppinen L, Hyvönen T, Jänne J, Eloranta TO (1990) Human spermidine synthase: cloning and primary structure. DNA Cell Biol 9(2):103–110
- Casero RA, Pegg AE (1993) Spermidine/spermine N1-acetyltransferase—the turning point in polyamine metabolism. FASEB J 7(8): 653–661
- Casero RA, Pegg AE (2009) Polyamine catabolism and disease. Biochem J 421(3):323–338
- Xie X, Gillies RJ, Gerner EW (1997) Characterization of a diamine exporter in Chinese hamster ovary cells and identification of specific polyamine substrates. J Biol Chem 272(33):20484–20489
- 21. Vujcic S, Liang P, Diegelman P, Kramer DL, Porter CW (2003) Genomic identification and biochemical characterization of the

mammalian polyamine oxidase involved in polyamine back-conversion. Biochem J 370(1):19–28

- Wu T, Yankovskaya V, McIntire WS (2003) Cloning, sequencing, and heterologous expression of the murine peroxisomal flavoprotein, N1-acetylated polyamine oxidase. J Biol Chem 278(23): 20514–20525
- Soulet D, Gagnon B, Rivest S, Audette M, Poulin R (2004) A fluorescent probe of polyamine transport accumulates into intracellular acidic vesicles via a two-step mechanism. J Biol Chem 279(47):49355–49366
- Belting M, Persson S, Fransson L-Å (1999) Proteoglycan involvement in polyamine uptake. Biochem J 338(2):317–323
- Belting M, Mani K, Jönsson M, Cheng F, Sandgren S, Jonsson S, Ding K, Delcros J-G, Fransson L-Å (2003) Glypican-1 is a vehicle for polyamine uptake in mammalian cells a pivotal role for nitrosothiol-derived nitric oxide. J Biol Chem 278(47):47181– 47189
- Wallace HM, Fraser AV, Hughes A (2003) A perspective of polyamine metabolism. Biochem J 376(1):1–14
- Palmer AJ, Wallace HM (2010) The polyamine transport system as a target for anticancer drug development. Amino Acids 38(2):415–422
- Byers TL, Pegg AE (1989) Properties and physiological function of the polyamine transport system. Am J Phys Cell Phys 257(3): C545–C553
- Delcros J-G, Tomasi S, Carrington S, Martin B, Renault J, Blagbrough IS, Uriac P (2002) Effect of spermine conjugation on the cytotoxicity and cellular transport of acridine. J Med Chem 45(23):5098–5111
- Mandel JL, Flintoff WF (1978) Isolation of mutant mammalian cells altered in polyamine transport. J Cell Physiol 97(3):335–343
- Corcé V, Renaud SP, Cannie I, Julienne K, Gouin SG, Loréal O, Gaboriau F, Deniaud D (2014) Synthesis and biological properties of Quilamines II, new iron chelators with antiproliferative activities. Bioconjug Chem 25(2):320–334
- Bergeron RJ, McManis JS, Weimar WR, Schreier K, Gao F, Wu Q, Ortiz-Ocasio J, Luchetta GR, Porter C, Vinson JT (1995) The role of charge in polyamine analog recognition. J Med Chem 38(13): 2278–2285
- Bergeron RJ, McManis JS, Liu CZ, Feng Y, Weimar WR, Luchetta GR, Wu Q, Ortiz-Ocasio J, Vinson JT (1994) Antiproliferative properties of polyamine analogs: a structure-activity study. J Med Chem 37(21):3464–3476
- Bergeron RJ, Neims AH, McManis JS, Hawthorne TR, Vinson JR, Bortell R, Ingeno MJ (1988) Synthetic polyamine analogs as antineoplastics. J Med Chem 31(6):1183–1190
- Bergeron RJ, McManis JS, Franklin AM, Yao H, Weimar WR (2003) Polyamine–Iron chelator conjugate. J Med Chem 46(25): 5478–5483
- Nick H, Acklin P, Lattmann R, Buehlmayer P, Hauffe S, Schupp J, Alberti D (2003) Development of tridentate iron chelators: from desferrithiocin to ICL670. Curr Med Chem 10(12):1065–1076
- Bergeron RJ, Singh S, Bharti N, Jiang Y (2010) Design, synthesis, and testing of polyamine vectored iron chelators. Synthesis 2010(21):3631–3636
- Bergeron RJ, Bharti N, Wiegand J, McManis JS, Yao H, Prokai L (2005) Polyamine-vectored iron chelators: the role of charge. J Med Chem 48(12):4120–4137
- Wang C, Delcros J-G, Biggerstaff J, Phanstiel IV O (2003) Molecular requirements for targeting the polyamine transport system. Synthesis and biological evaluation of polyamine–anthracene conjugates. J Med Chem 46(13):2672–2682
- 40. Wang C, Delcros J-G, Biggerstaff J, Phanstiel IV O (2003) Synthesis and biological evaluation of N-(anthracen-9ylmethyl)triamines as molecular recognition elements for the polyamine transporter. J Med Chem 46(13):2663–2671

- Gardner RA, Delcros J-G, Konate F, Breitbeil F, Martin B, Sigman M, Huang M, Phanstiel IV O (2004) N-substituent effects in the selective delivery of polyamine conjugates into cells containing active polyamine transporters. J Med Chem 47(24):6055–6069
- Padariya M, Kalathiya U, Baginski M (2015) Structural and dynamic changes adopted by EmrE, multidrug transporter protein studies by molecular dynamics simulation. Biochim Biophys Acta Biomembr 1848(10):2065–2074
- 43. Breitbeil III F, Kaur N, Delcros J-G, Martin B, Abboud KA, Phanstiel IV O (2006) Modeling the preferred shapes of polyamine transporter ligands and dihydromotuporamine-C mimics: shovel versus hoe. J Med Chem 49(8):2407–2416
- 44. Wang C, Delcros J-G, Cannon L, Konate F, Carias H, Biggerstaff J, Gardner RA, Phanstiel IV O (2003) Defining the molecular requirements for the selective delivery of polyamine conjugates into cells containing active polyamine transporters. J Med Chem 46(24): 5129–5138
- 45. Kaur N, Delcros J-G, Imran J, Khaled A, Chehtane M, Tschammer N, Martin B, Phanstiel Iv O (2008) A comparison of chloroambuciland xylene-containing polyamines leads to improved ligands for accessing the polyamine transport system. J Med Chem 51(5): 1393–1401

- 46. Kaur N, Delcros J-G, Archer J, Weagraff NZ, Martin B, Phanstiel Iv O (2008) Designing the polyamine pharmacophore: influence of Nsubstituents on the transport behavior of polyamine conjugates. J Med Chem 51(8):2551–2560
- 47. Muth A, Kamel J, Kaur N, Shicora AC, Ayene IS, Gilmour SK, Phanstiel IV O (2013) Development of polyamine transport ligands with improved metabolic stability and selectivity against specific human cancers. J Med Chem 56(14):5819–5828
- Phanstiel O, Kaur N, Delcros J-G (2007) Structure-activity investigations of polyamine-anthracene conjugates and their uptake via the polyamine transporter. Amino Acids 33(2):305–313
- 49. Corcé V, Morin E, Guihéneuf S, Renault E, Renaud S, Cannie I, Tripier R, Lima LM, Julienne K, Gouin SG (2012) Polyaminoquinoline iron chelators for vectorization of antiproliferative agents: design, synthesis, and validation. Bioconjug Chem 23(9):1952–1968
- Kaur N, Delcros J-G, Martin B, Phanstiel IV O (2005) Synthesis and biological evaluation of dihydromotuporamine derivatives in cells containing active polyamine transporters. J Med Chem 48(11): 3832–3839