Trace Determination of Iron in Real Waters and Fruit Juice Samples Using Rapid Method: Optimized Dispersive Liquid-Liquid Microextraction with Synthesized Nontoxic Chelating Agent

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Received: 3 December 2018 /Accepted: 4 February 2019 / Published online: 27 February 2019© Springer Science+Business Media, LLC, part of Springer Nature 2019

Abstract

The purpose of this research was to optimize a new method for preconcentration and determination of trace iron concentrations in aqueous solutions. For this purpose, a newly synthesized ligand, 3-(3-hydroxy-2-methyl-4-oxopyridin-1(4H)-yl) benzoic acid (3- OH-3-MOPBA), was used in the dispersive liquid-liquid microextraction (DLLME) method coupled with UV–vis spectroscopy. The experiments considering input variables of extractant volume, disperser volume, salt concentration, and pH were designed with the aid of central composite design (CCD). The results were analyzed using response surface methodology (RSM). The limit of detection (LOD) was found to be 4.0 µg L⁻¹ under the optimized conditions. A calibration curve with a good linearity (R^2 = 0.9986) was obtained over the concentration range of 15–800 μ g L⁻¹. The relative standard deviations (RSD) were found to be around 2.1% $(n = 7)$. The main advantages of the developed method are simple application, environment friendly, short time, and low cost which makes this method to be applied routinely for measuring iron in various water samples.

Keywords 3-OH-3-MOPBA . Iron . Preconcentration . Response surface methodology . UV–vis spectroscopy

Introduction

Undoubtedly, iron has an essential role in many metabolic functions in the human body. Fe is involved in a wide range of metabolic and synthetic pathways including DNA synthesis, oxygen transport and storage, mitochondrial respiration, and citric acid cycle [\[1](#page-9-0), [2](#page-9-0)]. Due to the important role of iron in the human body, its disorder (overload or deficiency of iron) causes a variety of diseases [\[3](#page-9-0)–[5](#page-9-0)]. Therefore, the determination of iron ions in trace amounts in variety of matrices, especially in aqueous samples, has been subject of many studies [\[6](#page-9-0)–[11\]](#page-9-0). Several analytical techniques such as flame atomic absorption spectrometry (FAAS) [[12](#page-9-0)], ICP-mass spectrometry (MS) [\[13\]](#page-9-0), inductively coupled plasma-optical emission spectrometry (ICP-OES) [\[14](#page-9-0)], chromatography [[15](#page-9-0)], and UV–vis spectrophotometry [[16](#page-9-0)–[18\]](#page-9-0) have been used for the speciation of iron.

Since various analytical techniques are unable to directly measure very low amounts of iron, many separation and preconcentration methods, including coprecipitation [\[19](#page-9-0)], solid phase extraction [[20\]](#page-9-0), electro-analytical methods [\[21\]](#page-9-0), and liquid-liquid extraction [\[22](#page-9-0)] were manipulated. One of the methods that is widely used for preconcentration and separation metal ions and organic compounds is the dispersive liquid-liquid microextraction method [\[23](#page-9-0)–[29\]](#page-9-0). There is a recent review article on the latest innovations and trends in liquid phase microextraction techniques [[30](#page-9-0)]. The dispersive liquid-liquid microextraction techniques are derived from three-component solvent system: homogeneous liquid-liquid extraction and cloud point extraction methods [[31](#page-9-0)]. It would definitely be said that the most important feature of this system is its solvent velocity. This feature is due to the high contact between the aqueous solvent and the organic solvent that causes the sample to be extracted in fractions of minutes from the aqueous phase to the organic phase [\[32\]](#page-10-0).

In the present study, a new method for the determination of iron in small amounts by dispersive liquid-liquid microextraction method coupled with UV–vis spectrophotometry was optimized.

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While most of the chelating agent applied to measure iron is toxic, the newly synthesized ligand, 3-(3-hydroxy-2-methyl-4 oxopyridin-1(4H)-yl) benzoic acid (3-OH-3-MOPBA) [[33\]](#page-10-0), in this study is non-toxic and environment friendly [\[34\]](#page-10-0). In the previous work, we used hydroxy pyridine derivatives as a chelating agent [\[35](#page-10-0)]. In comparison, the main advantages of the current study include a much better limit of detection (4 µg L⁻¹), the broader dynamic range (15–800 µg L⁻¹), optimal pH (4.6) in terms of adjustment for acetate buffer, more environment friendly because of using ethanol as a disperser solvent, and ultimately the most significant advantage is that these kinds of ligands have been confirmed to be selective of iron ions in aqueous matrices.

The use of multivariate optimization has been increased in recent analytical chemistry studies [[36](#page-10-0)–[39](#page-10-0)]. This optimization method not only allows simultaneous analysis of several parameters contemporaneously, but also increases measurement performance and speed of analysis and, most importantly, reduces the number of experiments that consequently results in saving time and money [\[40\]](#page-10-0). One of the most widely used statistical methods that is used for initial multi-variable optimization is the factorial design method [[41\]](#page-10-0). One of the most functionally used designs in surface response is central composite design (CCD) [[42](#page-10-0)]. The CCD is an experimental design, useful in response surface methodology (RSM), for building a second order (quadratic) model for the response variable without needing to use a complete three-level factorial experiment [\[43\]](#page-10-0). In this study, the effective parameters for extraction and determination of iron ions in the proposed DLLME method were optimized using CCD and RSM. The RSM explores the relationships between several explanatory variables and one or more response variables. The main idea of RSM is to use a sequence of designed experiments to obtain an optimal response [\[44](#page-10-0)]. The proposed optimal method is a selective method that can be used simply (because of combination with ultraviolet–visible spectroscopy) for routine determination of iron concentrations in aqueous solutions.

Experimental

Instrumentation

The absorbance of the solutions was determined by Shimadzu UV-160PC (Tokyo, Japan) spectrophotometer. The solutions were centrifuged with A Hettich EBA 200 (Andreas Hettich GmbH & Co. KG, 78532 Tuttlingen, Germany). Ultrapure water was prepared by a Milli-Q water purification system (Millipore S.A.S. 67120 Molsheim-France). The pH was measured by InoLab® pH meter (WTW; model 7110, Germany).

Software

MINITAB software was used for experimental design and statistical analysis (MINITAB Release 17.3.1, Minitab, Inc., Pine Hall Road, State College, PA, USA).

Chemical and Standard Solutions

Analytical grade $Fe(NH_4)(SO_4)_2.12 H_2O$ (Merck, Darmstadt, Germany) in 1.0 mol L^{-1} HNO3 was used to prepare the stock solution (Fe³⁺ (1000 mg L⁻¹)). Diluted working standard solutions were prepared every day from the stock solutions. Buffer acetate (mixing an appropriate amount of sodium acetate (analytical grade) and acetic acid solution (Merck, Darmstadt, Germany-analytical grade) (0.5 mol L^{-1})) were used to adjust the pH solutions (pH 2–8). Hydroxylamine hydrochloride solution (NH₂OH·HCl, analytical grade) 5% (w/v) was from Merck (Darmstadt, Germany) and prepared in a 100-mL volumetric flask by dissolving 5.0 g of the reagent in distilled water. Stock solution of 3-(3-hydroxy-2 methyl-4-oxopyridin-1(4H)-yl) benzoic acid (3-OH-3- MOPBA, analytical grade) (0.0045 mol L^{-1}) was made by dissolving an adequate amount of 3-OH-3-MOPBA in 10 mL of methanol (analytical grade). All organic and inorganic reagents used in this study were obtained from Merck (Darmstadt, Germany, analytical grade $(\geq 99.0\%)$).

Analytical Procedure

In this procedure, a mixture of 150 μL of ethanol, 600 μL of carbon tetrachloride, and 1 mL 4.5×10^{-3} mol L⁻¹ of the ligand (3-OH-3-MOPBA) was injected to 5 mL of a sample of water containing 80 μg L^{-1} of iron (including 0.7 mL of reducing agent (NH₂OH·HCl) 5% (w/v) and 1 mL of 0.5 mol L^{-1} acetate buffer (pH 4.5)). The effect of this injection is formation of a cloudy solution. Then, this cloudy solution is centrifuged at 4500 rpm for 3 min. Finally, the extraction phase was diluted in microcell 350 μL with ethanol, and the light absorption was read as absorbance at λ_{max} (546 nm) against a reagent blank.

Calculations

In order to evaluate the impact of independent factors on DLLME method, extraction recovery (R) and enrichment factor (EF) were calculated as follows:

$$
R\% = \frac{C_{sed} \times V_{sed}}{C_0 \times V_{aq}} \times 100 = EF \times \frac{V_{sed}}{V_{aq}} \times 100
$$
 (1)

$$
EF = \frac{C_{sed}}{C_0} \tag{2}
$$

Fig. 1 The absorption spectrums of the reagent blank (a) and the complex (b) after proposed method. Extraction conditions: 80 μg L⁻¹ of Fe³⁺, 1 mL of 45 \times 10^{-4} mol L⁻¹ of 3-OH-3-MOPBA, pH 4.5, 150 μL CCl4, 600 μL ethanol, 0.7 mL 5%- NH₂OH·HCL as a reducer agent, and 1 mL of 0.5 mol L^{-1} acetate buffer (pH 4.5)

where $R\%$ is the extraction recovery, V_{sed} is the volume of the sedimented phase, V_{aq} is the volume of the aqueous sample, and C_{sed} is the sedimented phase and was calculated from an external calibration graph obtained by conventional liquidliquid extraction followed by spectrophotometric determination of iron ions. C_0 represents the initial analyte concentration in the aqueous sample.

Results and Discussion

A dispersive liquid–liquid microextraction procedure based on the reaction between 3-OH-3-MOPBA as a chelating agent and Fe was developed for separation and determination of iron. To find the appropriate conditions for the proposed method, multiple parameters were investigated.

In this research, parameters such as selection of extraction and disperser solvents, volume of reducing agent, and chelating agent concentration by one variable at a time and parameters such as pH, extraction, and disperser volume were optimized by response surface modeling methods. Also, the effect of the added salt concentration was optimized by both one

variable at a time and response surface modeling methods, which both methods confirmed each other.

After all of the above-mentioned parameters were optimized, the extraction of iron with optimized parameters (analytical procedure) was performed.

One Variable at a Time Optimized

Selection of Wavelength

Figure 1 presents the results obtained from the analysis of the ligand and complex absorption spectra. From this graph, the two absorption bands at 546 and 662 nm could be observed (according to the extraction conditions presented in the analytical procedure): the absorption spectrums of the reagent blank (1 mL of 45×10^{-4} mol L⁻¹ of 3-OH-3-MOPBA) (a) and the complex (80 μg L⁻¹ of Fe³⁺, 1 mL of 45 \times 10⁻⁴ mol L⁻¹ of 3-OH-3-MOPBA, 150 μL CCl4, 600 μL ethanol 0.7 mL 5%-NH₂OH·HCL as a reducer agent and 1 mL of 0.5 mol L−¹ acetate buffer (pH 4.5)) after optimized DLLME technique (b).

Fig. 2 Influence of extraction and disperser solvents kind on the extraction recovery of iron obtain from proposed method. Extraction conditions: 100 μL CCl4, 500 μL ethanol, $80 \mu g L^{-1}$ of Fe³⁺, 1 mL of

 45×10^{-4} mol L⁻¹ of 3-OH-3-MOPBA, 0.7 mL 5%-NH₂OH·HCL as a reducer agent, and 1 mL of 0.5 mol L^{-1} acetate buffer (pH 4.5)

Table 1 Independent variables, their symbols, and levels for CCD and optimum conditions predicted

^a The predicted value of extraction recovery by CCD is 90.24%

The observed bands (546 and 662 nm) are attributed to the formation of Fe-(3-OH-3-MOPBA) complex. The

more intense band of 546 nm was selected for determination of iron concentration.

Table 2 The central composite design in various experimental and the responses

Selection of Organic Solvents

The choice of suitable organic solvents in the DLLME procedure is very important. In this study, various organic solvents including carbon tetrachloride, tetrachloroethylene, dichloromethane, and chloroform were used as extraction solvent. (experimental conditions 80 μg L⁻¹ of Fe³⁺, 1 mL of 45 \times 10^{-4} mol L⁻¹ of 3-OH-3-MOPBA, 0.7 mL 5%-NH₂OH· HCL as a reducer agent, 1 mL of 0.5 mol L^{-1} acetate buffer 150 μL and pH (4.5)). Also, organic disperser solvents such as acetone, acetonitrile, ethanol, and methanol were used. Figure [2](#page-2-0) shows the results of using different types of extraction and disperser solvents. The ethanol and methanol extraction efficiency are approximately the same. So, ethanol was selected because it is non-toxic. Due to the fact that the mixture of ethanol and carbon tetrachloride had the best extraction efficiency, they were chosen as a disperser and extractor solvents, respectively.

Influence of the NH₂OH·HCl Concentration

To evaluate the impact of volume of hydroxylamine hydrochloride) 5%-NH₂OH·HCL (on extraction recovery, various volumes (0.1–2 mL) of reducer agent were used, while other parameters were kept constant (experimental conditions: 80 μg L⁻¹ of Fe³⁺, 1 mL of 45×10^{-4} mol L⁻¹ of 3-OH-3-MOPBA, 1 mL of 0.5 mol L^{-1} acetate buffer, pH (4.5), 150 μL CCl₄, and 600 μL ethanol). At first, the extraction efficiency increased by increasing the volume of reducing agent to 0.7 mL. Then, a steady extraction efficiency was observed in volume of reducing agent from 0.7 to 0.9 mL. Finally, the increase in the volume of the reducing agent for higher than 0.9 mL resulted in a decrease in the extraction efficiency. Therefore, the volume of 0.7 mL was chosen as the optimum volume of the reducing agent.

Fig. 3 Pareto chart of factorial screening experiment ($p < 0.05$). A, B, C, and D are pH, extraction solvent volume (EX), dispersive solvent volume (DIS), and salt addition $(w/v \%)$, respectively. Experimental conditions: 1 mL of 45 × 10−⁴ mol L−¹ of 3-OH-3- MOPBA, CCl₄ (extraction solvent), ethanol (disperser solvent), 0.7 mL 5%- NH₂OH·HCL as a reducer agent, NaCl (evaluation of ionic strength), and acetate buffer (to pH adjusted)

Influence of Salting-Out

The effect of ionic strength on the DLLME method was studied by applying different concentrations of sodium chloride $(0-5\%$ w/v), while other parameters were kept constant (experimental conditions: 80 μg L^{-1} of Fe³⁺, 1 mL of 45×10^{-4} mol L⁻¹ of 3-OH-3-MOPBA, 0.7 mL 5% -NH₂OH·HCL as a reducer agent, 1 mL of 0.5 mol L⁻¹ acetate buffer, pH (4.5), 150 μ L CCl₄, and 600 μ L ethanol). The results indicated that the salt addition does not affect the extraction efficiency. Therefore, all the experiments were done without increasing the salt amount. This issue has been reaffirmed with insignificant data in response surface methodology section (multi-variation optimized section).

Influence of 3-OH-3-MOPBA

The influence of chelating agent concentration on the recovery of iron ions was evaluated $(0.0001 -$ 0.008 mol L^{-1}), while other parameters were kept constant (experimental conditions: 80 μg L^{-1} of Fe³⁺, 0.7 mL 5%-NH2OH·HCL as a reducer agent, 1 mL of 0.5 mol L^{-1} acetate buffer, pH (4.5), 150 µL CCl₄, and 600 μL ethanol). The recovery increased by an increase of the 3-OH-3-MOPBA concentration from 0.0005 to 0.0045 mol L^{-1} , and then it remains constant. Therefore, the 4.5×10^{-3} mol L⁻¹ was selected as the optimal concentration of 3-OH-3-MOPBA.

Multi-variation Optimized

Response Surface Methodology and Selection of Optimum Conditions

In this section, after primitive optimal conditions (a number of parameters were obtained in the one variable at a time), four important factors of pH, salt concentration (NaCl), volume of extractant $(CCl₄)$, and disperser (ethanol) solvents which affect the pre-concentration and extraction of iron ions were optimized employing the twolevel full factorial by central composite design (CCD) [\[45\]](#page-10-0). All condition tests performed in this section are described in the analytical procedure section (80 μ g L⁻¹ of Fe³⁺, 1 mL of 45×10^{-4} mol L⁻¹ of 3-OH-3-MOPBA, 0.7 mL 5%-NH2OH·HCL as a reducer agent and acetate buffer (to pH adjusted)). The encoded and non-encoded values of experimental variables are specified in Table [1.](#page-3-0) The data from the 31 experiments (include 16 cube points, 7 center points in cube, and 8 axial points) designed by MINITAB software version 17.3.1 derived from the extraction of iron ions into DLLME method are shown in Table [2.](#page-3-0) [[46](#page-10-0)]. All the experiments performed in accordance with the conditions stated in the analytical procedure section. The mathematical relations between independent variables and response are a quadratic polynomial equation as follows:

 $Y = 83.48 - 4.03 A + 3.59 B - 10.97 C - 10 A^2 - 3.07 B^2 - 7.12 C^2$ (3)

Fig. 4 2D/3D contour plots and response surface models of extraction recovery percent versus of Ex (extraction solvent volume) and pH (a) Dis (dispersive solvent volume) and pH (b). Experimental conditions: 1 mL of 45×10^{-4} mol L⁻¹ of 3-OH-3-MOPBA, 0.7 mL 5%- NH2OH·HCL as a reducer agent, acetate buffer (to pH adjusted), NaCl (evaluation of ionic strength), CCl₄ (extraction solvent), and ethanol (disperser solvent)

where Y is the predicted average extraction recovery that depends on the three factors of pH, volume of extractant, and disperser solvents. A, B, and C are pH, extractant solvent volume (EX), and dispersion solvent volume (DIS), respectively.

Figure [3](#page-4-0) shows the Pareto chart generated in this experiment. It is clear that DIS, pH, and EX with the lowest p values respectively are the most significant factors. Also, the effect of quadratic terms of pH^2 , $Ex²$, and $Dis²$ is statistically significant on $R\%$, but

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none of the two-way interaction is not significant on R% in 95% confidence level. Figure 4 illustrates the contour plot and three-dimensional surface response of extraction recovery modeling for some of the important factors. These types of curves can well illustrate the interactions between the factors. According to Fig. 4a, in the pH range of 4 to 5, and the solvent volume of the extractor of 140 μ L, the extraction efficiency is maximized. Also, as shown in Fig. 4b, in the range of pH 4 to 5 and the solvent volume of the disperser, about

Fig. 4 (continued)

600 μL has the highest extraction amount. The optimal predicted points of the experiment conditions have been shown in the last column of Table [1.](#page-3-0) In order to evaluate the validity of the predicted model, three experiments were carried out in accordance with predicted optimal conditions, with an average extraction recovery of $87 \pm$ 1.73 that showed good agreement between the results of the experimental results and the predicted values.

Influence of Extraction Time

Extraction time is one of the major parameters affecting the extraction efficiency. In DLLME method, it was defined as an interval between the time of injection of the mixture (disperser + extraction solvent) and the time of starting of centrifugation. For evaluating this parameter, different extraction times (0.5 to 90 min at constant

experimental conditions: 80 μg L^{-1} of Fe³⁺, 1 mL of 45 × 10^{-4} mol L⁻¹ of 3-OH-3-MOPBA, 0.7 mL 5%-NH₂OH· HCL as a reducer agent, 1 mL of 0.5 mol L^{-1} acetate buffer, pH (4.5), 150 μL CCl₄, and 600 μL ethanol) were studied. The results indicated that the extraction time has no significant effect on the extraction efficiency. This could be due to the infinitely large surface area between the extraction solvent and the aqueous phase, susceptible to extracting the uranyl ions. Therefore, DLLME technique is very fast, and this is the most important advantage of it, as reported by other researchers. In this method, centrifuging of sample solution in the extraction procedure takes the most time which is about 3 min.

Influence of Foreign Ions

There is always a great problem with coexisting ions along with environmental samples to detect metallic ions in very low amounts using instrumental techniques [[47\]](#page-10-0). In order to investigate the interference of the concomitant ions, we used 5 mL of the sample containing iron ions (80 μ g L⁻¹) along with different amounts of other ions (experimental conditions: 1 mL of 45×10^{-4} mol L⁻¹ of 3-OH-3-MOPBA, 0.7 mL 5% -NH₂OH·HCL as a reducer agent, 1 mL of 0.5 mol L⁻¹ acetate buffer, pH (4.5), 150 μL CCl₄, and 600 μL ethanol). As the results in Table 3 show, at a tolerable concentration of concomitant ions, less than 5% deviation in absorbance reading was observed. Therefore, the presence of coexisting ions in different amounts did not affect our optimized method.

Analytical Method Validation

The analytical performance characteristics of the developed DLLME method were studied at optimal value of input variables. At optimized condition, the calibration graph with excellent linearity ($R^2 = 0.9986$) was observed for the iron concentration from 15 to 800 ng mL^{-1} . The mathematical equation of calibration curve was as $y =$ $0.0022x + 0.0585$ by the optimized DLLME technique (experimental conditions 80 μg L^{-1} of Fe³⁺, 1 mL of

Table 3 Interferences of foreign ions for determination of iron (80 μg L−¹)

Coexisting ions	Tolerance ratio (w/w)	
EDTA	50	
SCN^-	100	
Al^{3+}	300	
Cu^{2+} , Sn^{2+} , Ag ⁺ , Tartarate, Mn ²⁺ , Co^{2+} , Zn^{2+} , Cd^{2+} , Pb^{2+} , Hg^{2+}	500	
$CH_3COO^-, SO_4^{2-}, NO_3^-, Cr^{2+}, Co^{3+}$	800	
$Li+$, Na ⁺ , K ⁺ , Ca ²⁺ , Mg ²⁺	1000	

 45×10^{-4} mol L⁻¹ of 3-OH-3-MOPBA, 0.7 mL 5%-NH₂OH·HCL as a reducer agent, 1 mL of 0.5 mol L^{-1} acetate buffer, pH (4.5), 150 μL CCl₄, and 600 μL ethanol). The limit of detection was calculated according to $LOD = 3$ SB/m in SB and m standard deviation of the blank and the slope of the calibration graph, respectively. LOD was observed to be 4 μ g L⁻¹. To determine the relative standard deviation (RSD), iron concentration of 50 μg L^{-1} was measured ten times at optimal condition. RSD value was found to be 2.1%. The enrichment factor was 23 (8 mL of iron aqueous solution resulted in 350 μL).

The developed DLLME method was successfully tested for actual aqueous samples (spiked tap, well, and mineral waters). Real water samples were preconcentrated, and their iron concentrations were measured. The results of iron determination of real samples are shown in Table 4.

Comparison of the Proposed Method with Previous Study

The results of this study were compared with other methods by the same technique used in preconcentration and determination of iron ions. These results are shown in Table [5.](#page-8-0) In contrary to other methods, present method has a lower

Table 4 Analytical results for iron in various real water samples

Sample	Fe added $(\mu g/L)$	Fe found, mean \pm SD ^a $(\mu g/L)$	Recovery $(\%)$
Well water ^b		98.00 ± 0.47	
	40	138.1 ± 0.32	100
	80	172.1 ± 0.55	97
	100	202.4 ± 0.28	102
Tap water ^c		33.00 ± 00.61	
	40	67.29 ± 0.2	92
	80	111.1 ± 0.41	98
	100	125.7 ± 0.36	95
Mineral water ^d		Not detected	
	40	41.97 ± 0.83	105
	80	83.57 ± 0.27	104
	100	97.00 ± 0.35	97
Fruit juice ^e		5.000 ± 0.95	
	40	44.10 ± 0.95	98
	80	83.01 ± 0.95	98
	100	104.6 ± 0.95	100

^a Standard deviation $(n = 3)$

^b Collected from Isin Hormozgan Iran

c From drinking water of laboratory

^d Aquafina mineral water bought from market

e Bought from market

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 $\overline{1}$

detection limit (4 µg L^{-1}), lower RSD (2.1%), the broader dynamic range (15–800 μg L^{-1}), higher enrichment factor (23), and shorter extraction time (3 min) (experimental conditions 80 μg L⁻¹ of Fe³⁺, 1 mL of 45×10^{-4} mol L⁻¹ of 3-OH-3-MOPBA, 0.7 mL 5%-NH₂OH·HCL as a reducer agent, 1 mL of 0.5 mol L^{-1} acetate buffer, pH (4.5), 150 μL CCl₄, and 600 μL ethanol). Also, due to the use of non-toxic chelating agent, our method is environment friendly [[34](#page-10-0)]. According to the results, it can be said that our optimized DLLME method coupled with UV–vis spectrophotometer is a simpler and more selective method for iron determination in comparison with previously developed methods.

Conclusions

This paper describes an effective optimized method for the determination and preconcentration of iron ions in a variety of water samples. The important parameters were optimized using both one variable at a time and response surface modeling methods. In this study, parameters such as selection of extraction $(CCl₄)$ and disperser solvents (ethanol), volume of reducing agent $(0.7 \text{ mL } 5\% \text{-} NH_2OH \cdot HCL)$, and chelating agent concentration (1 mL of 45×10^{-4} mol L⁻¹ of 3-OH-3-MOPBA) by one variable at a time and parameters such as pH (4.5), extraction (150 μ L), and disperser (600 μ L) volume were optimized by response surface modeling methods. Also, the effect of the added salt concentration was optimized by both one variable at a time and response surface modeling methods, which both methods confirmed each other.

Because of the special selectivity between the chelating agent and the iron ions $(pH = 4.5)$, the extraction was performed with the least ion interference. The use of spectrophotometric instrumentation is one important merit of simplicity, cheapness, portability, and so on in comparison with other previous studies. The present method has lower detection limits and lower RSDs within less sample volume. Due to the low levels of organic solvents, as well as the non-toxicity of our non-toxic chelating agent, this method is considered as an environmentally friendly method. It is believed that the proposed procedure can be useful for analysis and monitoring of iron level in aqueous, environmental, and biological samples.

Compliance with Ethical Standards

Conflict of Interest The authors declare that they have no conflict of interest.

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References

- 1. Muckenthaler MU, Rivella S, Hentze MW, Galy B (2017) A red carpet for iron metabolism. Cell 168(3):344–361. [https://doi.org/10.](https://doi.org/10.1016/j.cell.2016.12.034) [1016/j.cell.2016.12.034](https://doi.org/10.1016/j.cell.2016.12.034)
- 2. Volani C, Doerrier C, Demetz E, Haschka D, Paglia G, Lavdas AA, Gnaiger E, Weiss G (2017) Dietary iron loading negatively affects liver mitochondrial function. Metallomics 9(11):1634–1644
- 3. Fibach E, Rachmilewitz EA (2017) Iron overload in hematological disorders. Presse Med 46(12, Part 2):e296–e305. [https://doi.org/10.](https://doi.org/10.1016/j.lpm.2017.10.007) [1016/j.lpm.2017.10.007](https://doi.org/10.1016/j.lpm.2017.10.007)
- 4. Percy L, Mansour D, Fraser I (2017) Iron deficiency and iron deficiency anaemia in women. Best Pract Res Clin Obstet Gynaecol 40: 55–67. <https://doi.org/10.1016/j.bpobgyn.2016.09.007>
- 5. Lozoff B, Georgieff MK (2006) Iron deficiency and brain development. In: Seminars in pediatric neurology, vol 3. Elsevier, Amsterdam, pp 158–165
- 6. Pepper SE, Borkowski M, Richmann MK, Reed DT (2010) Determination of ferrous and ferric iron in aqueous biological solutions. Anal Chim Acta 663(2):172–177. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.aca.2010.01.056) [aca.2010.01.056](https://doi.org/10.1016/j.aca.2010.01.056)
- Zhu Y, Pan D, Hu X, Han H, Lin M, Wang C (2017) An electrochemical sensor based on reduced graphene oxide/gold nanoparticles modified electrode for determination of iron in coastal waters. Sensors Actuators B Chem 243:1–7. [https://doi.org/10.1016/j.snb.](https://doi.org/10.1016/j.snb.2016.11.108) [2016.11.108](https://doi.org/10.1016/j.snb.2016.11.108)
- 8. Kassem MA, Amin AS (2013) Spectrophotometric determination of iron in environmental and food samples using solid phase extraction. Food Chem 141(3):1941–1946. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.foodchem.2013.05.038) [foodchem.2013.05.038](https://doi.org/10.1016/j.foodchem.2013.05.038)
- 9. Hoshino M, Yasui H, Sakurai H, Yamaguchi T, Fujita Y (2011) Improved spectrophotometric determination of total iron and iron (III) with o-hydroxyhydroquinonephthalein and their characterization. Yakugaku Zasshi 131(7):1095–1101
- 10. Sobhi HR, Azadikhah E, Behbahani M, Esrafili A, Ghambarian M (2018) Application of a surfactant-assisted dispersive liquid-liquid microextraction method along with central composite design for micro-volume based spectrophotometric determination of low level of Cr(VI) ions in aquatic samples. Spectrochim Acta A Mol Biomol Spectrosc 202:36–40. <https://doi.org/10.1016/j.saa.2018.05.031>
- 11. Rastegarzadeh S, Pourreza N, Larki A (2013) Dispersive liquid– liquid microextraction of thiram followed by microvolume UV–vis spectrophotometric determination. Spectrochim Acta A Mol Biomol Spectrosc 114:46–50. [https://doi.org/10.1016/j.saa.2013.](https://doi.org/10.1016/j.saa.2013.05.020) [05.020](https://doi.org/10.1016/j.saa.2013.05.020)
- 12. Rihana-Abdallah A, Li Z, Lanigan KC (2015) Cloud point extraction of Iron and its detection using flame atomic absorption spectrometry. In: Trace materials in air, soil, and water, vol 1210. ACS symposium series, vol 1210. American Chemical Society, pp 183– 193. [https://doi.org/10.1021/bk-2015-1210.ch00910.1021/bk-](https://doi.org/10.1021/bk-2015-1210.ch00910.1021/bk-2015-1210.ch009)[2015-1210.ch009](https://doi.org/10.1021/bk-2015-1210.ch00910.1021/bk-2015-1210.ch009)
- 13. De Jong J, Schoemann V, Lannuzel D, Tison J-L, Mattielli N (2008) High-accuracy determination of iron in seawater by isotope dilution multiple collector inductively coupled plasma mass spectrometry (ID-MC-ICP-MS) using nitrilotriacetic acid chelating resin for pre-concentration and matrix separation. Anal Chim Acta 623(2): 126–139
- 14. Moss JC, Hardaway CJ, Richert JC, Sneddon J (2010) Determination of cadmium copper, iron, nickel, lead and zinc in crawfish [Procambrus clarkii] by inductively coupled plasma optical emission spectrometry: a study over the 2009 season in Southwest Louisiana. Microchem J 95(1):5–10. [https://doi.org/](https://doi.org/10.1016/j.microc.2009.07.008) [10.1016/j.microc.2009.07.008](https://doi.org/10.1016/j.microc.2009.07.008)
- 15. Ichinoki S, Fujita S, Fujii Y (2008) Selective determination of iron ion in tap water by solvent extraction with 3, 4-dihydro-3-hydroxy-

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4-oxo−1, 2, 3-benzotriazine, followed by reversed phase HPLC. J Liq Chromatogr Relat Technol 32(2):281–292

- 16. Frag EY, Abd El-Ghany NA, Fattah MAEL (2018) Physicochemical properties and characterization of iron (II) electrochemical sensor based on carbon paste electrode modified with novel antimicrobial Carboxymethyl chitosan-graft-poly(1 cyanoethanoyl-4-acryloyl-thiosemcarbazide) copolymers. J Electroanal Chem 808:266–277. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jelechem.2017.12.018) [jelechem.2017.12.018](https://doi.org/10.1016/j.jelechem.2017.12.018)
- 17. de Oliveira e Silva AF, de Castro WV, de Andrade FP (2018) Development of spectrophotometric method for iron determination in fortified wheat and maize flours. Food Chem 242:205–210. <https://doi.org/10.1016/j.foodchem.2017.08.110>
- 18. Solana-Altabella A, Sánchez-Iranzo MH, Bueso-Bordils JI, Lahuerta-Zamora L, Mellado-Romero AM (2018) Computer vision-based analytical chemistry applied to determining iron in commercial pharmaceutical formulations. Talanta 188:349–355. <https://doi.org/10.1016/j.talanta.2018.06.008>
- 19. Grotti M, Soggia F, Ardini F, Frache R (2009) Determination of sub-nanomolar levels of iron in sea-water using reaction cell inductively coupled plasma mass spectrometry after Mg (OH) 2 coprecipitation. J Anal At Spectrom 24(4):522–527
- 20. Chen Y, Feng S, Huang Y, Yuan D (2015) Redox speciation analysis of dissolved iron in estuarine and coastal waters with on-line solid phase extraction and graphite furnace atomic absorption spectrometry detection. Talanta 137:25–30
- 21. Lu M, Rees NV, Kabakaev AS, Compton RG (2012) Determination of iron: electrochemical methods. Electroanalysis 24(8):1693–1702
- 22. da Silva GC, JWSDd C, Dweck J, Afonso JC (2008) Liquid–liquid extraction (LLE) of iron and titanium by bis-(2-ethyl-hexyl) phosphoric acid (D2EHPA). Miner Eng 21(5):416–419. [https://doi.org/](https://doi.org/10.1016/j.mineng.2007.10.013) [10.1016/j.mineng.2007.10.013](https://doi.org/10.1016/j.mineng.2007.10.013)
- 23. Faraji H, Helalizadeh M (2017) Lead quantification in urine samples of athletes by coupling DLLME with UV-vis spectrophotometry. Biol Trace Elem Res 176(2):258–269
- 24. Wang Y, Zhang J, Zhao B, Du X, Ma J, Li J (2011) Development of dispersive liquid–liquid microextraction based on solidification of floating organic drop for the determination of trace nickel. Biol Trace Elem Res 144(1–3):1381–1393
- 25. Khan S, Soylak M, Kazi TG (2013) Room temperature ionic liquidbased dispersive liquid phase microextraction for the separation/ preconcentration of trace Cd2+ as 1-(2-pyridylazo)-2-naphthol (PAN) complex from environmental and biological samples and determined by FAAS. Biol Trace Elem Res 156(1–3):49–55
- 26. Habibi H, Mohammadi A, Farhoodi M, Jazaeri S (2018) Application and optimization of microwave-assisted extraction and dispersive liquid–liquid microextraction followed by highperformance liquid chromatography for the determination of Oleuropein and Hydroxytyrosol in olive pomace. Food Anal Methods 11(11):3078–3088
- 27. Saraji M, Ghambari H (2018) Dispersive liquid–liquid microextraction based on liquid anion exchanger for the direct extraction of inorganic anions. J Chromatogr A 1574:27–35
- 28. Lima DL, Silva CP, Otero M (2018) Dispersive liquid-liquid microextraction for the quantification of venlafaxine in environmental waters. J Environ Manag 217:71–77
- 29. Yilmaz E, Soylak M (2013) Ionic liquid-linked dual magnetic microextraction of lead (II) from environmental samples prior to its micro-sampling flame atomic absorption spectrometric determination. Talanta 116:882–886
- 30. Yilmaz E, Soylak M (2016) Latest trends, green aspects, and innovations in liquid-phase–based microextraction techniques: a review. Turk J Chem 40(6):868–893
- 31. Shamsipur M, Fattahi N, Pirsaheb M, Sharafi K (2012) Simultaneous preconcentration and determination of 2, 4-D, alachlor and atrazine in aqueous samples using dispersive liquid–

liquid microextraction followed by high-performance liquid chromatography ultraviolet detection. J Sep Sci 35(20):2718–2724

- 32. Çabuk H, Akyüz M, Ata Ş (2012) A simple solvent collection technique for a dispersive liquid–liquid microextraction of parabens from aqueous samples using low-density organic solvent. J Sep Sci 35(19):2645–2652
- 33. Marczenko Z, Balcerzak M (2000) Separation, preconcentration and spectrophotometry in inorganic analysis, vol 10. Elsevier, Amsterdam
- 34. Mesquita RB, Suárez R, Cerdà V, Rangel M, Rangel AO (2013) Exploiting the use of 3, 4-HPO ligands as nontoxic reagents for the determination of iron in natural waters with a sequential injection approach. Talanta 108:38–45
- 35. Borzoei M, Zanjanchi MA, Sadeghi-aliabadi H, Saghaie L (2018) Optimization of a methodology for determination of iron concentration in aqueous samples using a newly synthesized chelating agent in dispersive liquid-liquid microextraction. Food Chem 264: 9–15. <https://doi.org/10.1016/j.foodchem.2018.04.135>
- 36. Campos AF, Cassella RJ (2018) Determination of acetate and formate in vegetable oils by ion chromatography after multivariate optimization of the extraction process using a Doehlert design. Food Chem 269:252–257. [https://doi.org/10.1016/j.foodchem.](https://doi.org/10.1016/j.foodchem.2018.06.136) [2018.06.136](https://doi.org/10.1016/j.foodchem.2018.06.136)
- 37. Ferreira SL, Lemos VA, de Carvalho VS, da Silva EG, Queiroz AF, Felix CS, da Silva DL, Dourado GB, Oliveira RV (2018) Multivariate optimization techniques in analytical chemistry-an overview. Microchem J 140:176–182
- 38. Vanini G, Souza MO, Carneiro MT, Filgueiras PR, Bruns RE, Romão W (2015) Multivariate optimisation of ICP OES instrumental parameters for Pb/Ba/Sb measurement in gunshot residues. Microchem J 120:58–63
- 39. Grijalba AC, Martinis EM, Wuilloud RG (2017) Inorganic selenium speciation analysis in Allium and Brassica vegetables by ionic liquid assisted liquid-liquid microextraction with multivariate optimization. Food Chem 219:102–108
- 40. Arabi M, Ghaedi M, Ostovan A (2017) Synthesis and application of in-situ molecularly imprinted silica monolithic in pipette-tip solidphase microextraction for the separation and determination of gallic acid in orange juice samples. J Chromatogr B 1048:102–110
- 41. Soylak M, Narin I, de Almeida Bezerra M, Ferreira SLC (2005) Factorial design in the optimization of preconcentration procedure for lead determination by FAAS. Talanta 65(4):895–899
- 42. Hao J, Wang F, Wang X, Zhang D, Bi Y, Gao Y, Zhao X, Zhang Q (2012) Development and optimization of baicalin-loaded solid lipid nanoparticles prepared by coacervation method using central composite design. Eur J Pharm Sci 47(2):497–505
- 43. Y-j LIU, R-w XIANG (2007) Application of central composite design/response surface methodology in pharmacy experiment design [J]. Chin J Modern Appl Pharm 6:007
- 44. Bezerra MA, Santelli RE, Oliveira EP, Villar LS, Escaleira LA (2008) Response surface methodology (RSM) as a tool for optimization in analytical chemistry. Talanta 76(5):965–977
- 45. Ryan TP, Morgan J (2007) Modern experimental design. J Stat Theory Pract 1(3–4):501–506
- 46. Myers RH, Montgomery DC, Vining GG, Borror CM, Kowalski SM (2004) Response surface methodology: a retrospective and literature survey. J Qual Technol 36(1):53–77
- 47. Zhao S, Chen F, Zhang J, Ren S, Liang H, Li S (2015) On-line flame AAS determination of traces Cd (II) and Pb (II) in water samples using thiol-functionalized SBA-15 as solid phase extractant. J Ind Eng Chem 27:362–367
- 48. Rasouli Z, Hassanzadeh Z, Ghavami R (2016) Application of a new version of ga-rbf neural network for simultaneous spectrophotometric determination of Zn (II), Fe (II), Co (II) And Cu (II) in real samples: an exploratory study of their complexation abilities toward mtb. Talanta 160:86–98
- 49. Rojas FS, Ojeda CB, Pavón JC (2012) Determination of iron by dispersive liquid-liquid microextraction procedure in environmental samples. Am J Chem 2(1):28–32
- Silva SG, Oliveira PV, Rocha FR (2010) A green analytical procedure for determination of copper and iron in plant materials after cloud point extraction. J Braz Chem Soc 21(2):234–239
- 51. Pehlivan E, Kara D (2007) Iron speciation by solid phase extraction and flame atomic absorption spectrometry using N, N′-bis-(2-hydroxy-5-bromobenzyl)-2-hydroxy-1, 3-diiminopropane. Microchim Acta 158(1–2):137–144
- 52. Ondigo D, Tshentu Z, Torto N (2013) Electrospun nanofiber based colorimetric probe for rapid detection of Fe2+ in water. Anal Chim Acta 804:228–234